

Coenzyme-Catalyzed Cleavage of Cobalt-Carbon Bonds in the
Oxidation of cis-Dialkylcobalt(III) Complexes by Oxygen

Shunichi FUKUZUMI,* Tatsushi GOTO, Kunio ISHIKAWA, and Toshio TANAKA
Department of Applied Chemistry, Faculty of Engineering,
Osaka University, Suita, Osaka 565

Redox coenzyme analogues (riboflavin, aminopterin, and lumazine) catalyze the cleavage of cobalt-carbon bonds in the oxidation of cis-dialkylcobalt(III) complexes, $\text{cis-}[\text{R}_2\text{Co}(\text{bpy})_2]\text{ClO}_4$ ($\text{R} = \text{PhCH}_2, \text{C}_2\text{H}_5, \text{CH}_3$; $\text{bpy} = 2,2'$ -bipyridine) by oxygen in the presence of HClO_4 in acetonitrile.

The cleavage of the cobalt-carbon bond of methylcobalamin is believed to be a key step in a vitamin B_{12} dependent methyl transfer reaction, catalyzed by tetrahydrofolate methyltransferase which contains redox coenzymes, i.e., reduced forms of pterin and flavin coenzymes,¹⁾ although the exact mechanism has not been established. Thus, considerable interest has been focused on the cleavage of cobalt-carbon bonds in organocobalt complexes.²⁻⁶⁾ However, no coenzyme-catalyzed cleavage of cobalt-carbon bonds has so far been reported. We report herein the first example of coenzyme-catalyzed cleavage of cobalt-carbon bonds in the oxidation of cis-dialkylcobalt(III) complexes by oxygen. The redox coenzymes used as catalysts in this study are a flavin (riboflavin) and pterin coenzyme analogues (aminopterin and lumazine).

The cis-dialkylcobalt(III) complexes, $\text{cis-}[\text{R}_2\text{Co}(\text{bpy})_2]\text{ClO}_4$ ($\text{R} = \text{PhCH}_2, \text{C}_2\text{H}_5, \text{CH}_3$; $\text{bpy} = 2,2'$ -bipyridine), are stable towards oxygen in acetonitrile at 298 K. However, $\text{cis-}[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$ reacts with oxygen in the presence of a strong acid such as HClO_4 in acetonitrile to produce benzyl hydroperoxide which decomposes to yield benzaldehyde as the final product (Table 1).⁷⁾ In the absence of oxygen, the cobalt-carbon bond was cleaved by the electrophilic attack of proton to yield toluene (Table 1). When a catalytic amount of riboflavin is added to an oxygen saturated acetonitrile solution of $\text{cis-}[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$ containing HClO_4 , the oxidation rate to yield benzaldehyde was enhanced significantly. The oxidation rate, monitored by the decay of the absorption band due to $\text{cis-}[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$, obeyed the pseudo-first-order kinetics. The observed rate constant increased linearly with increasing the catalyst concentration. Aminopterin and lumazine also exhibit the catalytic effect in the oxidation of $\text{cis-}[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$ by oxygen in the presence of HClO_4 (0.10 mol dm^{-3}).

The one-electron oxidation of $\text{cis-}[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$ by a strong one-electron oxidant, $[\text{Fe}(\text{phen})_3]^{3+}$ ($\text{Phen} = 1,10$ -phenanthroline), in the presence of oxygen gives the same product as the case of the coenzyme-catalyzed oxidation, i.e., benzaldehyde (Table 1).⁸⁾ In the absence of oxygen, however, the one-electron

Table 1. Yields of the products for catalytic (or noncatalytic) cleavage of the cobalt-carbon bonds of $\text{cis-}[\text{R}_2\text{Co}(\text{bpy})_2]^+$ by oxidants in the absence and presence of HClO_4 in acetonitrile (0.6 cm^3)

$\text{cis-}[\text{R}_2\text{Co}(\text{bpy})_2]\text{ClO}_4$ (10^{-5} mol)	Oxidant (10^{-5} mol)	HClO_4 (10^{-5} mol)	Product (%), based on the cobalt complex ^a)
R = PhCH_2	O_2 Riboflavin		
	2.5 6.9 0.10	6.0	PhCHO (100) PhCH_2OOH (trace)
	2.5 6.9 0	6.0	PhCHO (100) PhCH_2OOH (trace)
	2.0 0 0	6.0	PhCH_3 (100)
	O_2 $[\text{Fe}(\text{phen})_3]^{3+}$		
	2.4 6.9 94	0	PhCHO (99) PhCH_2OOH (trace)
	2.4 0 94	0	$\text{PhC}_2\text{H}_4\text{Ph}$ (98)
R = C_2H_5	O_2 Riboflavin		
	2.4 6.9 0.28	6.8 ^b)	C_4H_{10} (100) C_2H_6 (trace)
	2.1 6.9 0	20	C_4H_{10} (76) C_2H_6 (24) C_2H_4 (13)
	2.1 0 0	6.3	C_4H_{10} (25) C_2H_6 (75) C_2H_4 (41)
	O_2 $[\text{Fe}(\text{phen})_3]^{3+}$		
	2.0 6.9 94	0	C_4H_{10} (100) C_2H_6 (trace)
	2.0 6.9 0	0	C_4H_{10} (100) C_2H_6 (trace)
R = CH_3	O_2 Riboflavin		
	3.6 6.9 3.6	12	C_2H_6 (89) CH_4 (11)
	2.1 6.9 0	4.1	C_2H_6 (trace) CH_4 (100)
	2.1 0 0	4.1	C_2H_6 (trace) CH_4 (100)

a) Determined by ^1H NMR and GLC. b) In the presence of $5.4 \times 10^{-4} \text{ mol H}_2\text{O}$.

oxidation results in the cleavage of both cobalt-benzyl bonds to yield the coupling product, 1,2-diphenylethane (Table 1).⁹⁾ In the case of $\text{cis-}[(\text{C}_2\text{H}_5)_2\text{Co}(\text{bpy})_2]^+$, the riboflavin-catalyzed oxidation by oxygen in the presence of HClO_4 gives exclusively the coupling product, butane (Table 1). Without riboflavin, the cobalt-ethyl bond is slowly cleaved by HClO_4 to yield butane, ethane and ethylene in both the absence and presence of oxygen. The yield of coupling product, butane, is larger in the presence of oxygen, compared with the case in the absence of oxygen (Table 1). The one-electron oxidation by $[\text{Fe}(\text{phen})_3]^{3+}$ in both the absence and presence of oxygen also gives the coupling product, butane, in contrast with the case of $\text{cis-}[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$ (Table 1).⁴⁾ In the case of $\text{cis-}[(\text{CH}_3)_2\text{Co}(\text{bpy})_2]^+$ as well, the riboflavin-catalyzed oxidation gives the coupling product, ethane, but small amount of methane is also formed because of the facile electrophilic cleavage of the cobalt-carbon bond by HClO_4 (Table 1). Without riboflavin, the cobalt-methyl bond is slowly cleaved to yield exclusively methane (Table 1).

Without HClO_4 , the redox coenzyme analogue (riboflavin, aminopterin, or lumazine) showed no catalytic activity. The effects of HClO_4 on the second-order rate constants for the cleavage of the cobalt-carbon bonds by the redox coenzyme analogues are shown in Fig. 1, together with the pseudo-first-order rate constants

for the electrophilic cleavage of the cobalt-carbon bonds in the absence of the redox coenzyme analogue for comparison. The $\log k_{\text{obsd}}$ values increase linearly with increasing the $\log[\text{HClO}_4]$ value, except for the $\text{cis}-[(\text{C}_2\text{H}_5)_2\text{Co}(\text{bpy})_2]^+$ -aminopterin system (Fig. 1). In the present systems, each coenzyme is protonated in the presence of HClO_4 in MeCN. Thus, upon the one-electron reduction, riboflavin semiquinone (FlH^\bullet) and lumazine semiquinone (LH^\bullet) may be further protonated to produce dihydroriboflavin radical cation ($\text{FlH}_2^{+\bullet}$) and dihydrolumazine radical cation ($\text{LH}_2^{+\bullet}$), respectively. Such protonation upon one-electron reduction is known to cause the positive shift of the one-electron reduction potential and thus, accelerates the electron transfer reaction.¹⁰⁾ In the case of aminopterin (P), the protonation of PH^\bullet may occur at high HClO_4 concentrations ($> 1 \text{ mol dm}^{-3}$) and

thus, the rate constant is constant with increasing the HClO_4 concentration up to 1 mol dm^{-3} (Fig. 1). In fact, the formation of dihydroriboflavin radical cation in the reaction of $\text{cis}-[(\text{C}_2\text{H}_5)_2\text{Co}(\text{bpy})_2]^+$ with riboflavin in the presence of HClO_4 under a degassed condition was confirmed by the ESR spectrum ($g = 2.0026$).¹¹⁾ In the case of the reaction with aminopterin, no radical cation has been detected.

Based on the above results, the catalytic mechanisms are shown in Schemes 1 and 2, where the protonated riboflavin (FlH^+) can be replaced by the protonated lumazine (LH^+) or aminopterin (P). In Scheme 1, the electron transfer from $\text{cis}-[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^+$ to FlH^+ occurs in the presence of HClO_4 to produce $\text{cis}-[(\text{PhCH}_2)_2\text{Co}(\text{bpy})_2]^{2+}$ and $\text{FlH}_2^{+\bullet}$. The cobalt-benzyl bond in the dibenzylcobalt(IV) complex may be readily cleaved to give benzyl radical, followed by the radical trap by oxygen to produce benzyl peroxy radical which then gives benzyl hydroperoxide by the abstraction of hydrogen atom from $\text{FlH}_2^{+\bullet}$, accompanied by regeneration of FlH^+ . The benzyl hydroperoxide decomposes to give the final product, benzaldehyde.

In the case of $\text{cis}-[\text{R}_2\text{Co}(\text{bpy})_2]^+$ ($\text{R} = \text{CH}_3$ and C_2H_5), the alkyl radical formed upon the one-electron oxidation by FlH^+ may undergo the facile coupling reaction with $[\text{RCo}(\text{bpy})_2]^{2+}$ to yield the coupling product R-R before the radical trap by oxygen (Scheme 2). The dihydroriboflavin radical cation $\text{FlH}_2^{+\bullet}$ may be further reduced by $[\text{Co}(\text{bpy})_2]^{2+}$ in the presence of HClO_4 to produce FlH_3^+ which is known to

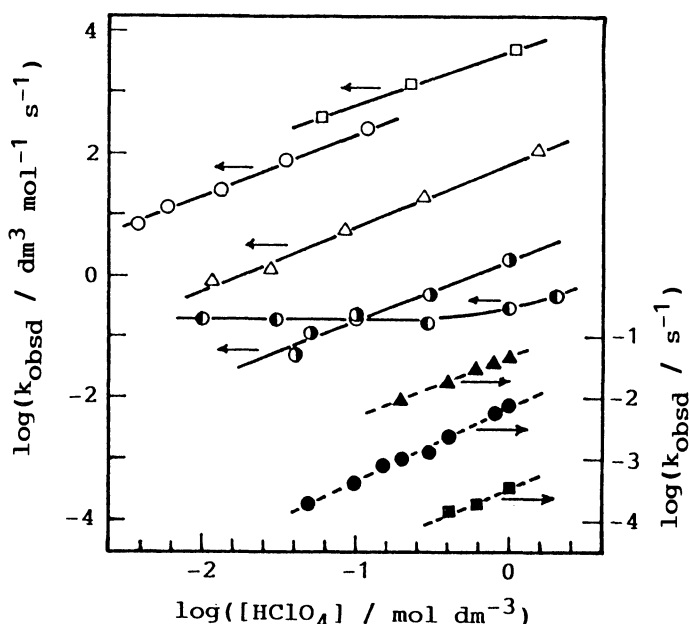
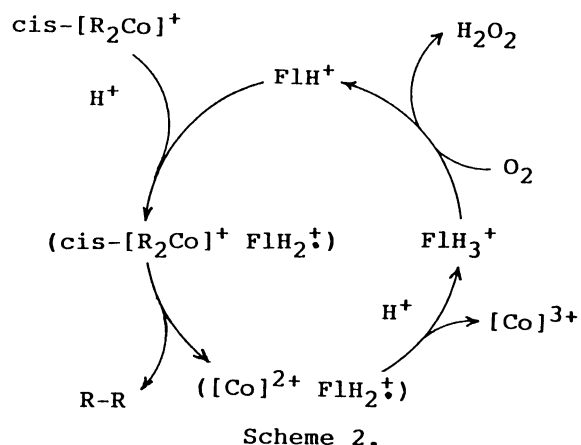
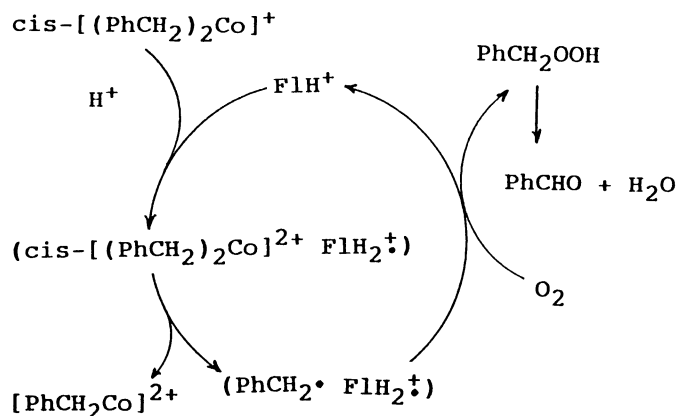


Fig. 1. Plots of $\log k_{\text{obsd}}$ vs. $\log[\text{HClO}_4]$ for the reactions of $\text{cis}-[\text{R}_2\text{Co}(\text{bpy})_2]^+$ [$\text{R} = \text{PhCH}_2$ (\square), C_2H_5 (\circ), CH_3 (\triangle)] with riboflavin, for the reaction of $\text{cis}-[(\text{C}_2\text{H}_5)_2\text{Co}(\text{bpy})_2]^+$ with aminopterin (\bullet) and lumazine (\circ) in the presence of HClO_4 , and for the reactions of $\text{cis}-[\text{R}_2\text{Co}(\text{bpy})_2]^+$ [$\text{R} = \text{CH}_3$ (\blacktriangle), C_2H_5 (\bullet), PhCH_2 (\blacksquare)] with HClO_4 in MeCN at 298 K.



be readily oxidized by oxygen to regenerate FlH^+ .¹²⁾ The mechanistic difference between Schemes 1 and 2 may be ascribed to the difference in the reactivity between the alkyl radicals; benzyl radical is much more stable than methyl or ethyl radical in the coupling reaction with $[\text{RCo}(\text{bpy})_2]^{2+}$.⁷⁾

References

- 1) R. T. Traylor, "B₁₂," ed by D. Dolphin, Wiley, New York (1982), Vol. 2.
- 2) J. Halpern, *Acc. Chem. Res.*, **15**, 238 (1982); R. H. Abeles and D. Dolphin, *ibid.*, **9**, 114 (1976); J. Halpern, S.-H. Kim, and T. W. Leung, *J. Am. Chem. Soc.*, **106**, 8317 (1984); **107**, 2199 (1985).
- 3) Y.-T. Fanchiang, *Organometallics*, **4**, 1515 (1985); *J. Chem. Soc., Dalton Trans.*, **1985**, 1375; *Inorg. Chem.*, **23**, 3428, 3983 (1984); **21**, 2344 (1982).
- 4) W. H. Tambllyn, R. J. Klingler, W. S. Hwang, and J. K. Kochi, *J. Am. Chem. Soc.*, **103**, 3161 (1981).
- 5) Y. Murakami, Y. Hisaeda, T. Ozaki, T. Tashiro, T. Ohno, Y. Tani, and Y. Matsuda, *Bull. Chem. Soc. Jpn.*, **60**, 311 (1987); Y. Murakami, Y. Hisaeda, T. Ozaki, and Y. Matsuda, *Chem. Lett.*, **1988**, 469.
- 6) S. Fukuzumi, K. Ishikawa, and T. Tanaka, *Organometallics*, **6**, 358 (1987); K. Ishikawa, S. Fukuzumi, and T. Tanaka, *Bull. Chem. Soc. Jpn.*, **60**, 563 (1987).
- 7) S. Fukuzumi, K. Ishikawa, and T. Tanaka, *Chem. Lett.*, **1986**, 1.
- 8) The decomposition of PhCH_2COOH to PhCHO may not occur by a homolytic pathway since no appreciable amount of PhCH_2OH has been detected.
- 9) S. Fukuzumi, K. Ishikawa, and T. Tanaka, *J. Chem. Soc., Dalton Trans.*, **1985**, 899; *Nippon Kagaku Kaishi*, **1985**, 62.
- 10) S. Fukuzumi, K. Ishikawa, K. Hironaka, and T. Tanaka, *J. Chem. Soc., Perkin Trans. 2*, **1987**, 751.
- 11) The hyperfine coupling constants of FlH_2^+ , determined by the computer simulation, are $a_5(\text{N}) = 0.585$, $a_5(\text{H}) = 1.17$, $a_8(\text{CH}_3) = 0.236$, $a_{10}(\text{N}) = 0.428$, $a_{10}(\text{CH}_2) = 0.431$ mT.
- 12) S. Fukuzumi, S. Kuroda, and T. Tanaka, *J. Am. Chem. Soc.*, **107**, 3020 (1985); S. Fukuzumi and T. Tanaka, "Photoinduced Electron Transfer," ed by F. A. Fox and M. Chanon, Elsevier, Amsterdam (1988), Chap. 11, in press.

(Received August 24, 1988)